

EVALUATION OF HAEMOLYTIC ACTIVITY OF SOME *CANDIDA* **SPECIES**

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Abstract

The ability to produce enzymes such as hemolysins is an important virulence factor for the genus Candida the objective of this study was to compare the hemolytic activity between *C. albicans* and other *Candida* species. A total of (100) *Candida* isolates representing (4) species are examine for their respective responses to an in hemolytic test. All strains of *Candida* specie isolated from the oral cavity of patients in Jumhuri Hospital in Kirkuk city. The four species isolated were: *C. albicans*, *C. glabra*, *C. krusei* and *C. kefyr*. Production of Hemolysin was evaluated on sabouraud dextrose agar containing chloramphenicol blood and glucose. A loop full of pure *Candida* culture was spot-inoculated and incubated for 24 hours at 37°C. Hemolytic activity was defined as the formation around the colonies of a translucent halo. 10 strains of *C. albicans* four strains of *C. glabra* 43 strains of *C. krusei* and 1 strain if *C. kefyr* subsequently that were studied produced hemolysins. Over all this study showed that hemolytic activity was detected in all isolates with minor differences seen between them the highest hemolytic activity detected in *C. krusei* (20-35mm) followed by *C. albicans* produces (25-33mm) while *C. glabra* (19-31mm) and the lowest hemolytic activity was detected in *C. kefyr* (14-16mm).

Key words : Candida sp., Haemolytic activity, Vaginal infection.

Introduction

Candidiasis is a multiple fungal disease that includes mucosal-cutanous visceral and proliferated infections caused by Candida species. Candida infection is one of the most common human mycoses (Edward et al., 2015). Candida species are the 3th to 4th most common bloodstream isolates in hospitalized patients with neutropenia or immunocompromised mainly from intensive, care units, (ICUs) (Pfaller et al., 2007). There is a high propagation of the mucosal-cutaneous forms particularly vaginal infections. The second most common vaginal infection is vaginitis caused by Candida species (Sobel, 2007). Candida species can produce a various of hydrolytic enzymes including proteases esterase lipases phosphatases and phospholipases (Cutle, 1991; Odds, 1998; Samaranayake and MacFarlane, 1990). These enzymes have received a great deal of attention in the past since they are known to interfere to candida pathogenesis especially by facilitates the hyphal incursion particularly seen in disseminated candidiasis (Fallon et al., 1997). While some of these hydrolytic enzymes such as phospholipases proteases and lipases have been explored (Hube et al., 1991; Lee, 1999; Tsang et al., 2007). The hemolytic activity shown by different Candida species is not well known (Manns and Mosser, 1994), prescribed an elegant yet simple plate assay method for observation the hemolytic activity of Candida albicans this method have been modified to estimate the hemolytic activity of different Candida species obtained from a variety of clinical manifold Candida species from a variety of clinical sources and to compare the species specific differences in the production of hemolysin qualitatively and quantitatively. (Watanabe et al., 1997) reported that Candida albicans excretes a hemolytic factor that causes hemoglobin to be released and is then used by the organism as an iron source. (Luo et al., 2001) reported that many species of Candida have two different types of hemolysins, alpha and beta hemolysin of which the nature is not yet understood. While many studies have been conducted on some of hydrolytic enzymes and hemolysin production in human isolates (Koga-Ito et al., 2006; Furlaneto-Maia et al., 2008). Research on famous

virulence factors particularly hemolytic activity offered by various animal-isolated *Candida* species and their products is limited Presently identifying virulence factors can play a key role in limiting pathogenesis of candidiasis and introducing new therapeutic agents (Ghannoum, 2000). Reviews have recounted that *Candida* spp. can secrete a number of exoenzymes such as hemolysin esterase proteinase and phospholipase needed for colonization and invasion host organs (Rudek, 1978; Watanabe *et al.*, 1999; Pakshir *et al.*, 2013).

Materials and Methods

Sampling and identification of yeasts

We studied (100) Candida isolated from the vaginal infection used in this study (4) species of Candida were detected. The species of these isolates were identified by vitik 2 system from May 2014 to April 2015, and the ability of these isolates to form germ tube. Colony characteristics on culture white to cream with characteristic yeast odor it grew rapidly and matured in 3 days (Mohammed and AL-Ahmadey, 2013). All *Candida* sp. strains isolation cultivation and preservation by (Kwon-Chung and Benett, 1992) on Sabouraud Dextrose Agar (SDA), they were sub-cultured on CHROM agar after the yeast colonies developed at 37°C for 48 hr. to evaluate the purity of the culture and colour



Fig. 1: (a) *C. albicans* grown on sabourauds agar (SDA) and CHROMagar. (b) Microscopy examination of *Candida albicans* showing buding and yeast cells.



Fig. 2: (a) *C. glabrata* grown on sabourauds (SDA) agar and CHROMagar. (b) Show mixed infections with more than one species of *Candida*.



Fig. 3: Germ tube of *Candida albicans* after 3 hr incubation.

of the colonies. This medium includes chromogenic substances (Staniszewska *et al.*, 2012). The method is based on the release of chromogenic breakdown products from different substrates by *candida* spp (Baker. 1967).

Identification using Vitik2 System

Vitik2 System has tested *Candida* isolates were tested by the reagent cards have (64) wells each with an individual test substratum. A suspension of each isolate was inoculated onto can 2 chromogenic agar plates at least twice before the testing (bioMérieux, France) and

> onto Sabouraud dextrose agar slants to ensure the purity and the viability of the cultures. As measured using a DensiChek instrument (bioMérieux), the inoculum suspensions for the VITEK 2 were prepared in sterile saline at a turbidity equal to a 2.0 McFarland standard. The individual test cards were automatically filled with the prepared culture suspension, sealed and incubated by the VITEK 2 instrument. The cards were incubated for 18 h at 35.5 °C and optical density readings were taken every (15) minutes automatically. The final results of the profile were compared with the database and the unknown organism the was identified.

Germ tube test

*Candida albicans*¹ ability of to form a germ tube was tested using Baker¹s protocol which was used to identify isolates. A single colony of cells was inoculated in human serum and incubated at 37C for 2-4 hr and then examined under the microscope for detected the germ tube. Both positive and negative germ tube





Fig. 4: (a,b,c,d) show the heamolytic activety of *C. albicans, C. glabrata C. krusei* and *C. kefyr* respectively.

production were classified as the isolates (Baker, 1967).

Hemolysin activity

Evaluated the production of hemolysin using methods with some modifications (Manns and Mosser, 1994). A loop full culture of pure Candida was inoculated into SDA medium containing chloramphenicol and incubated, for 24 hours at 37°C. 10⁸ cells/mL of suspension growth was prepare in sterile phosphate buffered (0.1 M, pH 7.2) saline using a spectrophotometer An aliquot (10 μ L) of the standardized suspension was cultured on to blood agar with glucose this medium was prepared with (5 mL) fresh blood per (10² mL) SDA supplemented with chloramphenicol and (3%) glucose. The pH was adjusted to (5.6 ± 0.2) Plates were incubated for 48 hr at 37°C. Colony and halo diameters were measured using a ruler and hemolytic activity was expressed in terms of the colony's diameter ratio of the to the degradation zone's outer diameter. The assay was performed in quadruplicate for each yeast isolate tested on two separate occasions. The halo zone for each isolates shown in Fig. 4.

Statistical analysis

Statistical analysis was conducted using SPSS for concentration human- depended T-test and compared mean by using Duncan multiple ranges under the level p-value ≤ 0.05 , p-value ≤ 0.01 was considered as statistically significant (Al- rawi, 2000).

Result and Discussion

Heamolytic activity of Candida species

(10) Strains of C. albicans, (4) of C. glabrata, (3) of C. krusei and (1) of C. kefvr species showed beta hemolysis blood SDA at 48 hours of incubation (Table 1). The quantitative data showed that C. albicans, C. glabrata and C. krusei's hemolytic activities were significantly higher than C. kefyr (p<0.01), apart from, there were no signifi-cant differences intraspecies in the β -hemolytic activities between these isolates C. albicans, C. glabrata and C. krusei (Table 1). As putative virulence factors hemolysins are known to contribute to Candida pathogenesis in particular to facilitate hyphal incursion candidasis speard (Luo et al., 2004). The hemolytic activities of yeast such as Candida genera has been investigated. (Lineraset al., 2007) reported a complementary hemolysis induced by C. albicans. (Watanabe et al., 1997) reported

that Candida albicans excretes a hemolytic factor that causes hemoglobin to be released and is then used by the organism as an iron source. (Luo et al., 2001) has stadied (80) isolates of (14) species of *Candida*, these authors reported that alpha and beta hemolysis had shown by C. albicans and others. Recently, we reported these species of Candida including C. albicans, C. tropi-calis C. glabrata, C. kefyr as well as C. krusei, have varying skills to produce hemolysins on human rabbit and SDA enriched with blood supplemented by glucose medium (Yigit and Aktas, 2009). There are limited Studies of *Candia*'s hemolysin activities isolated from oral isolates, carried out he first study of hemolytic activity from an isolated oral cavity of C. albicans (Tsang et al., 2007) reported that the hemolytic activity of an oral C. albicans isolated with type 2 diabetes mellitus patients was substantially higher than those controlled (a hemolysis index of 0.764±0.08 in the non-diabetic group vs. 0.673 ± 0.06 in the diabetic group). In this study Candida species isolated from vaginal infection were investigated in vitro hemolytic activities. C. albi-cans (n=10), C. glabrata (n=4), C. krusei (n=3) and C. kefyr (n=1) species exhibited β -hemolysis on blood SDA (Table 1). The quantitative data showed that the β hemolytic activity -of C. albicans (25-33mm), C. glabrata (19-31mm) and C. krusei (20-35mm) showed significantly higher beta-hemolytic activities than C. kefyr (14-16) (p<0.01) (Table 1). Furthermore, there were no

Table 1: Hemolysin activity species of Candida sp.

| Haemolysin activity mean±SD | Number of isolates | Species |
|--------------------------------|--------------------|------------|
| 25-33 (±2.538) | 10 | C.albicans |
| 20-35(±5.92) | 4 | C. glabra |
| 19-31 (± 7.94) | 3 | C. krusei |
| 14-16 (± 1.000) | 1 | C. kyfer |

significant intra species differences in the beta-hemolytic activities between isolates C. albicans, C. glabrata and C. krusei (Table 1). It is still necessary to consider the possibility that species specific hemolysis may texist. These hemolysis may vary of molecules and therefore have different rates of diffusion (Luo et al., 2001; Luo et al., 2004). The ability of pathogeni organisms to acquire elemental iron has been shown to be of crucial importance for their survival and the ability to infect the mammalian host (Weinberg, 1978; Bullen, 1981). Because there is essentially no free iron in the human host most pathogens acquire iron - indirectly containing compounds like hemoglobin (Belanger et al., 1995). To do so, However, the pathogen should be equipped with a mechanism that destroys the movement of the heme and allows the extraction of the elemental iron. The enzymes that mediate this activity are widely referred as hemolysins.

References

- Al- rawi, K.M. (2000). Introduction to statistical. Books house for printing and spreading, Al-Musol University.
- Baker. "Handbook of Bacteriological Technique". 2nd Editition, Butterworth Co. Ltd., London, 1967; 415-421.
- Belanger, M., C. Begin and M. Jacques (1995). Lipopolysaccharides of Actinobacillus pleuropneumoniae bind pig hemoglobin. *Infect. Immun.*, 63: 656-662.
- Bullen, J.J. (1981). The significance of iron in infection. *Rev. Infect. Dis.*, **3:** 1127-1138.
- Cutle, J.E. (1991). Putative virulence factors of *Candida albicans. Annu. Rev. Microbiol.*, **45**:187-218.
- Edward, J.E. (2015). Candida species. In Principles and Practice of Infectious Diseases, 8th ed.; J.E. Bennett, R. Dolin and M.J. Blaser, Eds.; Elsevier: Amsterdam, The Netherlands, 2879-2894.
- Fallon, K., K. Bausch, J. Noonan, E. Huguenel and P. Tamburini (1997). Role of aspartic proteases in disseminated *Candida albicans* infection in mice. *Infect. Immun.*, 65: 551-556.
- Furlaneto-Maia, L., A.F. Specian, F.C. Bizerra, M.T. de Oliveira and M.C. Furlaneto (2008). In vitro evaluation of putative virulence attributes of oral isolates of Candida spp. Obtained from elderly healthy individuals. *Mycopathologia*, **166**: 209-17.

Ghannoum, M.A. (2000). Potential role of phospholipases in

virulence and fungal pathogenesis. *Clin. Microbiol. Rev.*, **13(1):** 122-43.

- Hube, B., C.J. Turver, F.C. Odds, H. Eiffert, GJ. Boulnois, H. Kochel and R. Ruchel (1991). Identification, cloning and characterization of the gene for the secretory aspartate protease of *Candida albicans*. *Mycoses.*, **34(1)**: 59-61.
- Koga-Ito, C.Y., J.P. Lyon, V. Vidotto and M.A. de Resende (2006). Virulence factors and antifungal susceptibility of Candida albicans isolates from oral candidosis and control individuals. *Mycopathologia*, **161**: 209-217.
- Kwon-chung, K.J. and J.E. benentt (1992). Medical Mycology. Williams and Wilkins,105-161.
- Lee, G.C., S.J. Tang, K.H. Sun and J.F. Shaw (1999). Analysis of the gene family encoding lipases in *Candida rugosa* by competitive reverse transcription-PCR. *Appl. Environ. Microbiol.*, 65: 3888-3895.
- Lineras, B.E.C., S.E. Loreto, P.C. Silveira, P. Pozatti, A.L. Scheid and M.J. Santurio *et al.* (2007). Enzymatic and hemolytic activities of Candida dubliniensis straibs. *Rev. Inst. Med. Trop. S Paulo*, **49**: 203-206.
- Luo, G., L.P. Samaranayake, B.P.K. Cheung and G. Tang (2004). Reverse transcriptase polymerase chain reaction (RT-PCR) detected of HPL gene expression in Candida glabrata and its possible role *in vitro* haemolysin production. *APMIS*, **112:** 283-290.
- Luo, G, L.P. Samaranayake and J.J.Y. Yau (2001). Candida species exhibit differential in vitro hemolytic activities. J. Clin. Microbiol., 39: 2971-4.
- Manns, J.M., D.M. Mosser and H.R. Buckley (1994). Production of a hemolytic factor by Candida albicans. *Infect. Immun.*, 62: 5154-5156.
- Mohammed, S.A. and Z.Z. Al- Ahmadey (2013). Biofilm formation and antifungal susceptibility of Candida isolated from various clinical speciments. *Brit. Microbiol. Res. J.*, 3(4): 590-601.
- Odds, F.C. (1998). Candida and candidosis: a review and bibliography, 2nd ed. 1998. Bailliere Tindall, London, United Kingdom.
- Pakshir, K., K. Zomorodian, M. Karamitalab, M. Jafari, H. Taraz and H. Ebrahimi (2013). Phospholipase, esterase and hemolytic activities of Candida spp. Isolated from onychomycosis and oral lichenplanus lesions. J. Mycol. Med., 23(2): 113-8.
- Pfaller, M.A. and D.J. Diekema (2007). Epidemology of invasive candidiasis: A persistent public health problem. *Clin. Microbiol. Rev.*, **20**: 133-163. [Cross Ref] [Pub Med].
- Rudek, W.A. (1978). Esterase activity in Candida species. J Clin Microbiol., **8(6):** 756-9.
- Samaranayake, L.P. and T.W. MacFarlane (1990). Oral candidosis. Wright, Bristol, United Kingdom.
- Sobel, J.D. (2007). Vulvovaginal candidosis. *Lancet*, **369**: 1961-1971.
- Staniszewska, M., M. Bondaryk, K. Siennicka, J. Pilat and M.

Schaller (2012). Role of aspartic proteinases in Candida albicans virulence. Part 1. Substrate specificity of aspartic proteinases and Candida albicans pathogenasis. *J. Post.* **Microbiol.**, **51:2:** 127-135.

- Takahashi, M., Y. Banno and Y. ozawa (1991). Secreted Candida albicans phospholipases: purification and characterization of two forms of lysophospholipase-transayclase. J. Med. Vet. Microbiol., 29: 193-204.
- Tsang, C.S.P., F.C.S. Chu, W.K. Leung, L.J. Jin, L.P. Samaranayake and S.C. Siu (2007). Phospholipase, proteinase and haemolytic activities of Candida albicans isolated from oral cavities of patients with type 2 diabetes mellitus. *J. Med. Microbiol.*, 56: 1393-8.
- Watanabe, T., M. Takano, M. Murakami, H. Tanaka, A. Matsuhisa and N. Nakao (1999). Characterization of a haemolytic factor from Candida albicans. *Microbiology*, 145(Pt 3): 689-94.
- Watanabe, T., H. Tanaka, N. Nakao, T. Mikami and T. Matsumoto (1997). Hemoglobin is utilized by Candida albicans in the hyphal form but not yeast form. Biochem. *Biophys. Res. Commun.*, **1232:** 350-3.
- Weinberg, E.D. (1978). Iron and infection. *Microbiol. Rev.*, **42**: 45-66.
- Yigit, N. and E. Aktas (2009). Comparison of the efficacy of different blood medium in determining the hemolytic activity of Candida species. J. Mycol. Med., 19: 110-5.